37	<b>.</b>		2 🔪		
AD-A265 839 PORT DOCK	<b>JMENTATION</b>				
	16. RESTRICTIVE MARKINGS		<del></del>		
	f	Y/AVAILABILITY O			
2b. DECLASSIFICATION/DOWNGRADING SCHEDULE		Approved for public release; distribution is unlimited			
4. PERFORMING ORGANIZATION REPORT NUMBER(S)  NMR 1 93-24	S. MONITORING	ORGANIZATION R	EPORT NUMBER	(S) (2) (S)	
6a. NAME OF PERFORMING ORGANIZATION Naval Medical Research (If applicable)	7a. NAME OF MONITORING ORGANIZATION Naval Medical Command				
6c. ADDRESS (City, State, and ZIP Code) 8901 Wisconsin Avenue Bethesda, MD 20889-5055	7b. ADDRESS (City, State, and ZIP Code) Department of the Navy Washington, DC 20372-5120				
8a. NAME OF FUNDING/SPONSORING ORGANIZATION Naval Medical (If applicable) Research & Development Comman	9. PROCUREMEN	T INSTRUMENT ID	ENTIFICATION N	UMBER	
8c. ADDRESS (City, State, and ZIP Code)	10. SOURCE OF F	UNDING NUMBER	is .		
8901 Wisconsin Avenue Bethesda, MD 20889-5044	PROGRAM ELEMENT NO.	PROJECT NO.	TASK NO.	WORK UNIT ACCESSION NO.	
beciledad, iib about per	62787A	3M162787	A870-AR1288	DN243536	
Inconclusive hepatitis C virus antibody results in African  12. PERSONAL AUTHOR(S) Hyams KC, Okoth FA, Tukei PM, Vall.  13a. TYPE OF REPORT 13b. TIME COVERED FROM TO		RT (Year, Month, L		<del> </del>	
16. SUPPLEMENTARY NOTATION Reprinted from: Journal of Infectious Diseases 1993; Vol	. 167 pp. 254-2	55			
	18. SUBJECT TERMS (Continue on reverse if necessary and identify by block number) viral diseases; hepatitis C; epidemiology; serology				
FIELD GROUP SUB-GROUP viral disease					
19. ABSTRACT (Continue on reverse if necessary and identify by block number)					
	D'ELE JUN 1	FIC ECTE 1993			
		U			
	Tai Access == co-	HOLTY CLASSICS	TION		
20. DISTRIBUTION/AVAILABILITY OF ABSTRACT  MUNCLASSIFIED/UNUMITED SAME AS RPT. DITIC USERS	21. ABSTRACT SEC		HOR		

DD FORM 1473, 84 MAR

22a. NAME OF RESPONSIBLE INDIVIDUAL Phyllis Blum. Librarian.

83 APR edition may be used until exhausted.
All other editions are obsolete.

SECURITY CLASSIFICATION OF THIS P
UNCLASSIFIED

22b. TELEPHONE (Include Area Code) 22c. OFFICE SYMBOL (301) 295-2188 MRL/NMRI

## CORRESPONDENCE

93 6 15 25 5

93-13564

## Inconclusive Hepatitis C Virus Antibody Results in African Sera

Colleagues—In Africa, the prevalence of hepatitis C virus antibody (anti-HCV) is low compared to the frequency of serologic markers of hepatitis B, but high levels of false-positive anti-HCV

Informed consent was obtained from study subjects or their parents or guardians, and the research guidelines of the US Naval Medical Research Unit No. 3 Committee for the Protection of Human Subjects were followed in the conduct of this research.

The opinions and assertions contained herein are the private ones of the authors and are not to be construed as official or reflecting the views of the US Department of the Navy or the Department of Defense.

Financial support: Naval Medical Research and Development Command, NMC, NCR, Work Unit no. 62770A.3M162770A870.AR122.

Reprints or correspondence: Dr. Craig Hyams, Naval Medical Research Institute, 12300 Washington Ave., Rockville, MD 20852.

The Journal of Infectious Diseases 1993;167:254-5 

© 1993 by The University of Chicago. All rights reserved. 
∪u22-1899/93/6701-0046\$01.00

ELISA results have been reported [1-3]. To investigate the causes of false-positive results, sera from 688 outpatients living on the eastern coast of Kenya were evaluated.

Sera and epidemiologic data were originally obtained in 1987 for a survey of hepatitis B and arboviral infections [4, 5]. Sera had been stored for 4 years at -70°C before testing for anti-HCV by ELISA using a second-generation test kit (Abbott). The mean ELISA optical density ratio (ODR = mean test optical density/cutoff optical density) was calculated as previously described [6]. All sera repeatedly reactive by ELISA were further tested with a second-generation immunoblot assay (RIBA HCV Test System; Chiron, Emeryville, CA). Samples containing sufficient volume were further tested with a dot immunobinding assay (Abbott MATRIX HCV), which detects antibody to recombinant proteins derived from putative core, NS3, and NS4 regions of the HCV genome and is interpreted as positive for anti-HCV when there is reactivity to two distinct HCV proteins. Only sera positive by both ELISA and either the immunoblot or dot immunobinding assays were considered to be positive for anti-HCV and to represent active infection. In addition, 42 samples were randomly selected and tested for *Plasmodium fal*ciparum antibody using an indirect immunofluorescent assay (IFA).

The mean age of the population was 21 years (range, 1–79); 80% were female. Sixty-two subjects (9.0%) had a history of acute hepatitis, and 55 (8.0%) had received a blood transfusion. Sera from 179 subjects (26.0%; ODR = 1.3) were repeatedly reactive by ELISA for anti-HCV; however, only 5 sera (0.7%; ODR = 1.8) were positive and 18 (2.6%; ODR = 1.9) indeterminate by RIBA-2. By dot immunobinding assay, 5 serum samples were positive (ODR = 2.2) and 33 inconclusive (ODR = 1.6) among 129 samples that could be tested. Of 4 RIBA-2-positive samples with sufficient volume for further testing. I was positive and 3 negative by Abbott MATRIX HCV; of 13 RIBA-2-indeterminate samples further tested, 2 were positive, 2 negative, and 9 inconclusive by Abbott MATRIX HCV.

Hepatitis B surface antigen was found in 43 subjects (6.3%) and antibody to hepatitis B surface antigen or antibody to hepatitis B core antigen in 359 (52.2%). By IFA, 21 subjects (3.1%) had antibody to dengue-2, 14 (2.0%) to chikungunya, 24 (3.5%) to Sindbis, 15 (2.2%) to West Nile, 16 (2.3%) to Rift Valley fever, 18 (2.6%) to dugbe, 8 (1.2%) to Ganjam, and 0 to Crimean-Congo hemorrhagic fever viruses. All 42 tested sera had antibody to *P. falciparum*, and 40% had a titer of > 1:1000.

None of the subjects positive for anti-HCV by immunoblot or dot immunobinding assay had ever had an acute episode of jaundice or had received a blood transfusion. Subjects positive by RIBA-2 (mean age, 33 years) or Abbott MATRIX HCV (mean age, 41 years) tended to be older than other subjects (mean age, 21 years), but there was no age trend for subjects with false-positive ELISA or inconclusive RIBA-2 and Abbott MATRIX HCV results. False positive ELISA results and inconclusive immunoblot or dot immunobinding results were not associated with the subjects' sex, history of blood transfusion or acute hepatitis, antigen and antibody markers of hepatitis B infection, arboviral antibodies, or malaria antibody.

These data suggest that hepatitis C infection is very infrequent in eastern Kenya; however, false-positive ELISA results and inconclusive confirmatory assay results are common but unrelated to flavivirus infection (dengue, West Nile) [7] and antibody to P. falciparum. False positivity and inconclusive reactions could be related to prolonged storage of samples, cross-reacting antibody to unknown antigens, or infection by HCV variants [8-10]

## Kenneth C. Hyams, F. A. Okoth, P. M. Tukei, David S. Vallari, John C. Morrill, Gary Long, Jaya Bansal, and Niel Constantine

US Naval Medical Research Institute, Bethesda, US Army Medical Research Institute of Infectious Diseases, Frederick, and Pathology Department, University of Maryland, Baltimore, Kenya Medical Research Institute, Nairohi, Abbott Diagnostics Division, Abbott Laboratories, Abbott Park, Illinois

## References

- Aceti A, Taliani G, Bac C, Sebastiani A, Anti-HCV false positivity in malaria. Lancet 1990;336:1442-3.
- Jackson JB, Guay L. Goldfarb J, et al. Hepatitis C virus antibody in HIV-1 infected Ugandan mothers [letter]. Lancet 1991;337:551
- Tibbs CJ, Palmer SJ, Coker R, et al. Prevalence of hepatitis C in tropical communities: the importance of confirmatory assays. J Med Virol 1991;34:143-7.
- Hyams KC, Okoth FA, Tukei PM, et al. Epidemiology of hepatitis B in Eastern Kenya. J Med Virol 1989;28:106–9.
- Morrill JC, Johnson BK, Hyams C, et al. Serological evidence of arboviral infections among humans of coastal Kenya. J Trop Med Hyg 1991;94:166–8.
- van der Poel CL, Reesink HW, Schaasberg W, et al. Infectivity of blood seropositive for hepatitis C virus antibodies. Lancet 1990;335:558-60.
- Houghton M, Richman K, Han J, et al. Hepatitis C virus (HCV): a relative of the pestiviruses and flaviviruses. In: Hollinger FB, Lemon SM, Margolis HS, eds. Viral hepatitis and liver disease. Baltimore: Williams & Wilkins, 1991:328-33.
- Ellis LA, Brown D, Conradie JD, et al. Regional prevalence of hepatitis C antibodies in South Africa: an analysis of fresh and stored serum. In: Hollinger FB, Lemon SM, Margolis HS, eds. Viral hepatitis and liver disease. Baltimore: Williams & Wilkins. 1991:445-7.
- Wong DC, Diwan AR, Rosen L, et al. Non-specificity of anti-HCV test for seroepidemiological analysis. Lancet 1990;336:750-1.
- Chan SW, Simmonds P, McOmish F, et al. Serological responses to infection with three different types of hepatitis C virus [letter]. Lancet 1991;338:1391.

Accession For			
NTIS	GRA&I	3	
DTIC 7	rab		
Unaun			
Justification			
	ibution labilit	y Codes	
Avail and/or			
Dist	Special		
A	20		